288. Amino-acids and Peptides. Part VIII.* Determination of D- and L-Glutamic Acid by the Isotope-dilution Method.

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A new method based on the isotope dilution principle has been developed for the determination of the total (D- + L)glutamic acid in mixtures of aminoacids. In control experiments with synthetic mixtures the glutamic acid found was 99.2-99.3% of the theoretical amount. The procedure has been applied to the analysis of gliadin.

Accurate determination of both enantiomorphs of an amino-acid in a protein hydrolysate is difficult. Procedures based on partition chromatography (e.g., Moore and Stein, Ann. N.Y. Acad. Sci., 1948, 49, 265) and on ion-exchange (e.g., idem, J. Biol. Chem., 1951, 192, 663) determine the total of both isomers (D and L), but so far their accuracy appears to be

* Part VII, J., 1952, 594.

rather less than is often desirable for investigations of protein structure. The isotopedilution principle, avoiding as it does the difficulty of quantitative isolation, is clearly attractive for this purpose. In its most usual application, the L-amino-acid is isolated and therefore this form only is assayed. A modification has already been used by which both enantiomorphs of glutamic acid have been determined (Graff, Rittenberg, and Foster, J. Biol. Chem., 1940, 133, 745; Wieland and Paul, Ber., 1944, 77, 34). Isotopic DLglutamic acid hydrochloride is added to the mixture to be analysed, from which pure Land DL-glutamic acid hydrochlorides are then isolated; from the isotopic dilution of these two samples, the amount of D(and L)-glutamic acid may be calculated. It is however sometimes difficult to isolate the pure racemate, since it is normally present in low concentration and has a higher solubility than the enantiomorphs, and it is then necessary to determine the composition of the mixture of isomers by measuring its optical rotation.

We have developed an alternative method for the determination of the total (D- and L-)glutamic acid, using the isotope-dilution principle. A known weight of L-[^{15}N]glutamic acid hydrochloride is added to the amino-acid mixture, which is then benzoylated, and the solution is treated with acetic anhydride at room temperature for 48 hours. Under these conditions, benzoylglutamic acid is racemised and after hydrolysis of the benzoyl derivatives a sample of DL-glutamic acid hydrochloride can readily be isolated in the normal way. From the isotopic dilution, the total glutamic acid originally present can be calculated directly. Loss of glutamic acid before racemisation is achieved will affect the determination only if optical isomers are lost differentially, whilst after racemisation such loss will cause no error. Control experiments showed that under the above conditions benzoyl-L-glutamic acid is racemised to the extent of 97.5%; calculation shows that no appreciable error is introduced by this incompletion. For example, if 5% of the glutamic acid is originally present as the D-form and if racemisation is 95% complete, the error is less than 0.03% of the total glutamic acid (if isotopic dilution is 1:10).

This procedure has been tested by the analysis of known mixtures of amino-acids, containing both D- and L-glutamic acid. The isolation and conversion of the samples into nitrogen, and the mass-spectrometric analyses (for which we are indebted to the Atomic Energy Research Establishment, Harwell) followed the procedure described in Part V (J., 1951, 3047). The results are shown in Table 1. The total glutamic acid

Total glutamic acid hydrochloride		L-[¹⁵N]Glutamic acid hydro- chloride added,		DL-Gh hyd i	utamic acid rochloride solated	Total glutamic acid hydrochloride	
Synthetic	present		Atoms-%	No. of	Atoms-%	fou	nd
mixture	(g.)	g.	excess ¹⁵ N	cryst.	excess ¹⁵ N	(9	%)
A	0.7308	0.1036	14.956	5	1.869		99.25
				6	1.869, 1.872		99.20
				7	1.865, 1.865		99.50
						Mean	99 ·3
B	0.7051	0.0998	14.956	5	1.864, 1.865		99 ·37
				6	1.863, 1.862		99 •48
				7	1.869, 1.866		99 ·19
						Mean	99·3
С	0.6450	0.1000	14.998	5	2·021, 2·028		99.35
				6	2.030		99.06
				7	2·024, 2·023		99 ·43
						Mean	99·3
D	0.6870	0.0892	14.998	5	1.737		99.09
				6	1.731, 1.734		99·40
				7	1.736, 1.738		99·13
						Mean	99·2
E	2.0052	0.3485	15.214	4	$2 \cdot 266$		99·34
				5	$2 \cdot 266, 2 \cdot 268$		99 ·26
				6	$2 \cdot 265$		99 ·3 5
						Mean	99.3

TABLE 1.

found in each experiment is $99\cdot 2-99\cdot 3\%$ of the theoretical amount. From the close agreement with the result of Experiment *E*, in which the L-glutamic acid hydrochloride used in this work was itself analysed by this method, it seems likely that the accuracy may be slightly greater than these figures suggest.

As in the determination of the L-isomer, it is important to establish the purity of the isolated sample. Differential solubility tests (Part III, J., 1951, 1279; Part IV, *ibid.*, p. 1745) will not detect the presence of a slight excess of one enantiomorph in the racemate, since the D- and the L-form dissolve independently, but they will detect other impurities in the normal manner. Optical-rotation measurements will then give satisfactory evidence of optical purity, when the absence of foreign contaminants has been established. It is however a great advantage of this procedure that for most applications even gross contamination of the racemate by the L-isomer has a negligible effect on the analysis. For example, if 5% of the glutamic acid is originally present as the D-form, racemisation is 95% complete, and the isotopic dilution is 1:10, it may readily be calculated that the presence in the isolated racemate of 5% of the L-isomer would result in an additional error of only 0.02%.

The total glutamic acid obtained on hydrolysis of a sample of gliadin (kindly supplied by Professor A. C. Chibnall) was determined by this method and the results are shown in Table 2. The mean value of 45.8 g. of dry, ash-free protein compares with the corresponding figure of 44.1 g. for the L-isomer from the same sample (Part V, *loc. cit.*), the difference

		TAI	BLE 2 .				
		L-[15N]Glutamic acid hydro- chloride added,		DL-Glutamic acid hydrochloride isolated		Total glutamic acid found,	
Gliadin, g. 1∙5176	g. 0·0598	Atoms-% excess ¹⁶ N 14·810	No. of cryst. 4 5 6	Atoms-% excess ¹⁵ N 1·061 1·062, 1·060 1·061, 1·060 M	G. per 100 g. of gliadin * 45.83 45.85 45.87 ean 45.85	N, % of protein-N 24·64 24·65 24·66 24·65	
1.3332	0.0728	14·810	4 5 6	1.435, 1.431 1.434, 1.436 1.434 M	$ \begin{array}{r} 45.76 \\ 45.70 \\ 45.73 \\ ean \ 45.73 \\ 1.1 \\ 45.8 \\ 1.1 \\ 45.8 \\ 1.1 \\ $	24.6024.5724.5924.5924.62	
	Gliadin, g. 1·5176 1·3332	L-[18N] acid chlorid Gliadin, g. g. 1.5176 0.0598 1.3332 0.0728	TA1 L-[¹⁶ N]Glutamic acid hydro- chloride added, Gliadin, Atoms-% g. g. excess ¹⁶ N 1.5176 0.0598 14.810 1.3332 0.0728 14.810 Mcan	TABLE 2. L-[16N]Glutamic DL-Glu acid hydro- hydr chloride added, is Gliadin, Atoms-% No. of g. g. excess 16N cryst. 1.5176 0.0598 14.810 4 6 1.3332 0.0728 14.810 4 5 6 5 6	$\begin{array}{c ccccc} TABLE \ 2. \\ & & L-[^{16}N]Glutamic & DL-Glutamic acid \\ & acid hydro- & hydrochloride \\ & chloride added, & isolated \\ \hline Gliadin, & Atoms-\% & No. of & Atoms-\% \\ g. & g. & excess ^{16}N & cryst. & excess ^{16}N \\ 1 \cdot 5176 & 0 \cdot 0598 & 14 \cdot 810 & 4 & 1 \cdot 061 \\ & 5 & 1 \cdot 062, 1 \cdot 060 \\ & 6 & 1 \cdot 061, 1 \cdot 060 \\ & & M \\ 1 \cdot 3332 & 0 \cdot 0728 & 14 \cdot 810 & 4 & 1 \cdot 435, 1 \cdot 431 \\ & 5 & 1 \cdot 434, 1 \cdot 436 \\ & 6 & 1 \cdot 434 \\ & & M \\ Mcan of experiments (i) and (ii) \\ \hline \end{array}$	$\begin{array}{c cccccc} TABLE \ 2. \\ & & L-[^{16}N]Glutamic & DL-Glutamic acid \\ acid hydro- & hydrochloride & Total gi \\ chloride added, & isolated & acid f \\ \hline Gliadin, & Atoms-\% & No. of & Atoms-\% & G. per 100 g. \\ g. & g. & excess ^{16}N & cryst. & excess ^{16}N & of gliadin * \\ 1.5176 & 0.0598 & 14.810 & 4 & 1.060 & 45.83 \\ & 5 & 1.062, 1.060 & 45.85 \\ & 6 & 1.061, 1.060 & 45.85 \\ \hline 1.3332 & 0.0728 & 14.810 & 4 & 1.435, 1.431 & 45.76 \\ & 5 & 1.434, 1.436 & 45.70 \\ & 6 & 1.434 & 45.73 \\ \hline Mean \ 45.85 \\ \hline Mean \ of experiments (i) and (ii) : & 45.8 \\ \hline \end{array}$	

* Calc. for dry, ash-free protein.

(1.7 g.) representing D-glutamic acid in the hydrolysate. This is remarkably close to the estimate (1.85 g.) based on optical-rotation measurements of fractions isolated by the gravimetric method by Chibnall and his colleagues (*J. Int. Soc. Leather Trades' Chem.*, 1946, 30, 11). This amount may of course arise from racemisation during hydrolysis, a point that is now under investigation.

EXPERIMENTAL

Determination of the Total Glutamic Acid in Synthetic Mixtures.—(A) A solution of L-glutamic acid hydrochloride (0.7186 g.), DL-glutamic acid hydrochloride (0.0122 g.), and L-[¹⁵N]glutamic acid hydrochloride (0.1036 g.) in water (80 c.c.) was made alkaline to phenolphthalein paper with 5N-sodium hydroxide. Sodium carbonate (3.27 g.) was added and the solution was cooled to 0—2°. Benzoyl chloride (4.8 c.c.) was added, and the reaction mixture stirred vigorously for $4\frac{1}{2}$ hours, after which the solution still gave a faintly positive ninhydrin test. Further additions of sodium carbonate (0.82 g.) and benzoyl chloride (1.1 c.c.) were made; stirring was continued for another 3 hours, a negative ninhydrin test being then obtained. When the solution had regained room temperature, acetic anhydride (60 c.c.) was added and the mixture was left overnight. A further volume of acetic anhydride (60 c.c.) was then added, and the solution left for 30 hours, after which it was evaporated to dryness below 50° under reduced pressure. The residual white solid was dissolved in water (40 c.c.), made acid to Congo-red with 5N-hydrochloric acid, and again evaporated to dryness in vacuo. The residue was extracted with dried acetone, and the extract filtered and evaporated to dryness under reduced pressure, leaving a white solid which was refluxed for $4\frac{1}{2}$ hours with 15% hydrochloric acid (45.5 c.c.). On cooling of the solution, benzoic acid separated and was removed by extraction with benzene. The aqueous layer was evaporated to dryness under reduced pressure; the residue was dissolved in a small volume of water, treated with charcoal, filtered, and concentrated to small bulk. The solution was saturated with hydrogen chloride at 0°. After being kept overnight in the cold, a crystalline mass separated and was filtered off, washed, and dried (0.55 g.); it had $[\alpha]_{1}^{16} + 0.45^{\circ}$ (c, 3.0 in 5% hydrochloric acid).

The crude DL-glutamic acid hydrochloride was recrystallised in all-glass apparatus, the first fraction which should contain any unchanged L-glutamic acid hydrochloride being rejected. The second fraction was filtered off through sintered-glass, washed, and recrystallised. Crystallisation was repeated 7 times and portions from each crystallisation were powdered and transferred to a vacuum-desiccator containing calcium chloride and sodium hydroxide pellets.

Differential-solubility tests for purity were carried out on the later fractions by the vapourpressure method described in Part III (J, 1951, 1279). After the fourth crystallisation the product had no observable rotation (c, 1.0 in 5% hydrochloric acid).

No. of crystallisations	4	5	6	7
Yield (based on crude wt.), %	67	53	45	42
V. p. difference (mm. of oil)	4	3	3	3

(B) A solution of L-glutamic acid hydrochloride (0.6939 g.), DL-glutamic acid hydrochloride (0.0112 g.), and L-[¹⁵N]glutamic acid hydrochloride (0.0998 g.) in water (75 c.c.) was benzoylated and racemised as described for synthetic mixtures A. DL-Glutamic acid hydrochloride (0.49 g.; $[\alpha]_{7}^{17} + 0.5^{\circ}$, c, 2.5 in 5% hydrochloric acid) was isolated and crystallised seven times in all, in the usual manner.

No. of crystallisations	5	6	7
Yield (based on crude wt.), %	66	58	49
V. p. difference (mm. of oil)	3	4	3

(C) The mixture contained L-glutamic acid hydrochloride (0.6345 g.), DL-glutamic acid hydrochloride (0.0105 g.), L-[¹⁵N]glutamic acid hydrochloride (0.1000 g.), L-aspartic acid hydrochloride (0.08 g.), and L-lysine hydrochloride (0.015 g.).

The mixture was benzoylated and racemised in the usual manner. On completion of the hydrolysis of the N-benzoyl-DL-glutamic acid with 15% hydrochloric acid and removal of the free benzoic acid, the aqueous layer was evaporated to dryness, redissolved in water (20 c.c.), made acid to Congo-red, and treated with a solution of ether-soluble phosphotungstic acid (3.5 g. in 2 c.c. of water) at 70° for the removal of basic amino-acids; the amino-dicarboxylic acids were extracted by using the anion-exchange resin Amberlite IR-4B (both procedures were as described in Part V, J, 1951, 3047). DL-Glutamic acid hydrochloride (0.45 g.; $[\alpha]_D^{21} + 0.75^\circ$, c, 2.0 in 5% hydrochloric acid) was finally isolated and crystallised 7 times from 5N-hydrochloric acid as described in the previous experiments.

No. of crystallisations	5	6	7
Yield (based on crude wt.), %	58	44	39
V. p. difference (mm. of oil)	4	3	

(D) A solution of the mixture containing L-glutamic acid hydrochloride (0.6743 g.), DLglutamic acid hydrochloride (0.0127 g.), L-[¹⁵N]glutamic acid hydrochloride (0.0892 g.), Laspartic acid hydrochloride (0.08 g.), L-lysine hydrochloride (0.01 g.), DL-alanine (0.03 g.), L-tyrosine (0.01 g.), and glycine (0.01 g.) was treated as described for mixture C, and DLglutamic acid hydrochloride (0.46 g.; $[\alpha]_D^{20} + 0.6^\circ, c, 3.0$ in 5% hydrochloric acid) was isolated and crystallised 7 times in all.

No. of crystallisations	5	6	7
Yield (based of crude wt.), %	60	52	45
V. p. difference (mm. of oil)	5	3	4

(E) L-Glutamic acid hydrochloride (2.0052 g.) and L-[15 N]glutamic acid hydrochloride (0.3485 g.) were dissolved in water (60 c.c.). The analytical procedure followed that for mixture A, yielding 1.5 g. of crude DL-glutamic acid hydrochloride, which was crystallised 6 times.

No. of crystallisations	4	5	6
Yield (based on crude wt.), %	58	52	45
V. p. difference (mm. of oil)	3	4	3
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The isolated samples from each experiment were converted into nitrogen by Rittenberg's method ("Preparation and Measurement of Isotopic Tracers," Edwards, Ann Arbor, Michigan, 1947). The mass-spectrometric determinations were carried out by Mr. Palmer and his colleagues at the Atomic Energy Research Establishment, Harwell. The ¹⁵N : ¹⁴N ratio was calculated from the relative intensity of the peaks at mass numbers 29 and 28, as in Part V (*loc. cit.*); corrections were similarly applied for air and carbon dioxide. The total glutamic acid hydrochloride in the original mixture (X) was calculated from the formula $X = A [(C_o/C) - 1]$, where A is the amount of L-[¹⁵N]glutamic acid hydrochloride added and C_o and C are the atoms-% excess of ¹⁵N in the added and isolated materials respectively. The results are shown in Table 1.

Determination of the Total Glutamic Acid in Gliadin.—The sample of gliadin had 17.69(9)% Kjeldahl-N, corrected for 9.8% of moisture and 0.86% of ash. It was from the same batch as that used in Part V (loc. cit.).

(i) Gliadin (1.5176 g.) was treated with concentrated hydrochloric acid (8.12 c.c.) at room temperature for 18 hours, complete dissolution occurring. Water (6.2 c.c.) was added and the solution was then boiled under reflux for 24 hours. L-[¹⁵N]Glutamic acid hydrochloride (0.0598 g.) was added and the heating was continued for a further hour. After repeated evaporation *in vacuo* at 50° to remove excess of acid, the product was benzoylated and then racemised in the manner described above. DL-Glutamic acid hydrochloride (0.5 g.; $[\alpha]_D^{20}$ + 0.75°, c, 2.0 in 5% hydrochloric acid) was isolated as previously and crystallised 6 times in all. After the fourth crystallisation the product had no observable rotation (c, 0.8 in 5% hydrochloric acid).

No. of crystallisations	4	5	6
Yield (based on crude wt.), %	65	56	49
V. p. difference (mm. of oil)	4	3	3

(ii) In a duplicate experiment with 1.3332 g. of gliadin, 0.0728 g. of L-[¹⁵N]glutamic acid hydrochloride was added; 0.48 g. of crude DL-glutamic acid hydrochloride was isolated and crystallised 6 times.

No. of crystallisations	4	5	6
Yield (based on crude wt.), %	52	47	39
V. p. difference (mm. of oil)	3	3	

The subsequent procedure and calculation were as described above for the synthetic mixtures. The results are shown in Table 2.

We thank the Department of Scientific and Industrial Research for continued financial support, the Royal Society for a grant for the purchase of materials, and the Director of the Atomic Energy Research Establishment for the mass-spectrometric assays.

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